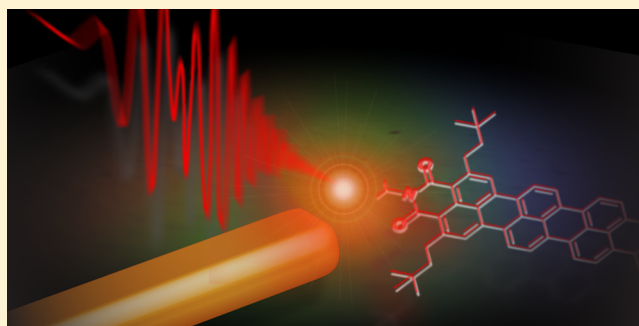


Ultrafast Meets Ultrasmall: Controlling Nanoantennas and Molecules

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ABSTRACT: We present a review on the advances of pulse control and ultrafast coherent excitation of both plasmonic nanoantennas and individual molecular systems, primarily based on the achievements in our group. Essential concepts from coherent control of ultrashort broadband laser pulses are combined with nanoscale diffraction limited detection and imaging of single photon emitters; that is, the central area of this work is where ultrafast meets ultrasmall. First, the critical role of dedicated pulse shaping and phase control is discussed, which is crucial to realize free of spatiotemporal coupling Fourier limited pulses inside a high numerical aperture microscope at the diffraction limited spot. Next we apply this scheme to plasmonic antennas, exploiting broadband two-photon excitation, to determine amplitude and phase of plasmonic resonances, to achieve ultrafast switching of nanoscale hotspots, and multicolor second harmonic detection for imaging applications. Subsequently, we address single molecules with phase-shaped pulses to control the electronic state population and retrieve single molecule vibrational dynamics response. We compare the response of a molecule to phase-locked with free phase multipulse excitation. Furthermore, we discuss phase control of excited state energy transfer in photosynthetic molecular complexes. Finally, we combine nanoscale plasmonics with single molecule detection to attain strong enhancement of both excitation and emission, with fluorescence lifetime shortening to the ps range. In conclusion, we anticipate that this review on ultrafast plasmonics and single emitter control will provide a useful view of the status of ultrafast nanophotonics and its application potential.

KEYWORDS: plasmonic nanoantenna, single molecule, coherent control, pulse-shaping, pump–probe, ultrafast nanoscopy



The experimental study of light–matter interactions at the level of individual nano-objects, to address fundamental spectroscopic properties beyond ensemble average, is now routine in many laboratories. Single molecules, semiconductor quantum dots (QDs), and metallic nanostructures are readily studied with a wide variety of optical techniques. In parallel, the field of ultrafast lasers has also developed strongly in the past two decades. Today, broadband lasers capable of delivering 15 fs pulses are used by many research groups for the study of the ultrafast processes and coherent dynamics in a large variety of systems. Bridging these two fields, the ultrasmall and ultrafast, is the current challenge in nanophotonics. The capacity to investigate femtosecond dynamics on the single nanoparticle level is promising to disentangle dynamical processes such as the intraparticle energy redistribution and interparticle energy transfer, which govern the ultrafast dynamics immediately after the absorption of a photon; or to examine the electronic dephasing and coherent processes associated with resonances in single molecules, QDs, and plasmonic nanoparticles.

Even though both single molecule detection^{1–5} and femtosecond spectroscopy in the condensed phase^{6–10} were thriving already in the '90s, it was not until after the year 2000 that the two fields gradually merged together and the first ultrafast microscopy of individual QDs¹¹ and molecules¹² was realized. The field of ultrafast nanophotonics is therefore quite

recent, and many research groups are currently developing new experimental strategies in this direction.^{11,13–25}

Our group has been active in the past 10 years on the ultrafast and coherent investigation of single molecules^{12,26–32} and nanostructures, with particular focus on plasmonic nanoantennas.^{28,33–36} Our objective has not only been to understand, but also to actively control ultrafast dynamics in individual nanoparticles. With the faster-than-ever progress in nanophotonics and nanotechnology and our growing ability to manipulate nanoscale dynamics, we might one day gain full control of the essential intra- and interparticle dynamical processes at the single nanoparticle level.

In experiments, the study of ultrafast light–matter interactions essentially comes down to delivering a specific series of ultrashort laser pulses with defined properties to the desired sample area and detecting relevant responses (observables) of the sample. One draws conclusions about the sample dynamics by manipulating one (or a few at the same time) of the accessible parameters of the ultrashort pulses: intensity, phase-time relation (chirp, pulse duration), color

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(and bandwidth), pulse sequence or polarization. Each of the laser pulse handles gives access to particular information: excitation power dependence (I), spectral position of resonances and relative excitation efficiency (λ), dynamics (Δt), anisotropy and mode directionality (polarization, P), and finally, temporal resolution/coherent effects (intrapulse and interpulse spectral phase, φ and ϕ). The aim is to eventually be able to actively induce the desired change in the sample in a controlled way. An analogy to puppetry can be drawn here (see Figure 1). In the learning stages, one pulls one string

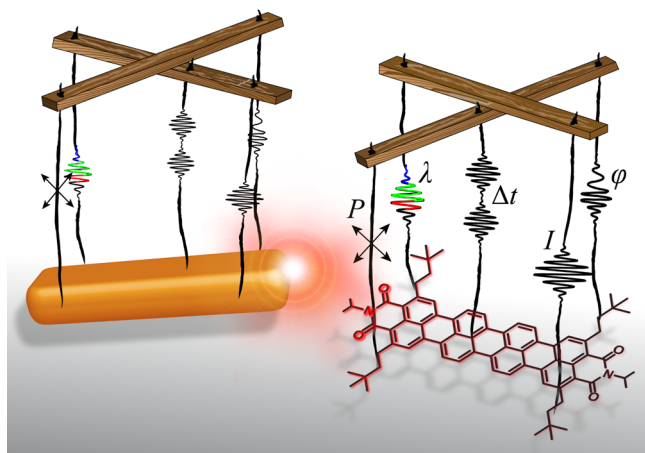


Figure 1. Puppetry vision of ultrafast coherent nanoscopy. By pulling the coherent control strings, the dynamics of nanoantennas, molecules, and their interactions can be orchestrated.

(parameter) at a time and looks at the response of the puppet. Once knowledge of which string leads to what movement is acquired, one knows exactly in what harmony, order, direction, and strength to pull the strings to obtain the desired movement, for instance, to make the puppet dance.

In a simple picture, the ultrafast response of a molecule or a nanoparticle to a photon excitation can be divided into two parts, the coherent and the incoherent regimes, which pertain to different time scales and therefore have different experimental requirements. In the coherent regime, the nanoparticle is sensitive to the phase properties of the excitation field. A properly designed pulse sequence, with tailored phase relation can therefore be used to control the response of the system, a concept that is usually named *coherent control*.^{37–42} In molecules at room temperature, however, the coherence is very short-lived, typically on the order of few 10s of femtoseconds, due to a large number of (coupled) degrees of freedom and strong coupling to the environment. Hence, accessing coherent dynamics requires ultrashort phase-controlled pulses. After the coherent regime, the system relaxes incoherently toward the equilibrium. The initially created population in the electronic excited states relaxes down the vibrational ladder to the lower lying vibrational states, a process termed (in case of molecules) intramolecular vibrational relaxation (IVR).⁴³ This typically occurs on a picosecond time scale. The particle then returns from the lowest excited state to the ground state through spontaneous emission within nanoseconds (with a probability given by the internal quantum yield).^{1,44} In this regime the particle is insensitive to the phase of the laser pulse with which it interacts.

In this review we summarize our efforts in understanding and manipulating these different ultrafast regimes in both single

molecules and plasmonic nanoantennas. The practical realization of these experiments is challenging, and the spread of ultrafast and ultrasmall studies throughout various research groups depends on the availability of a well-controlled experimental approach. In this review we therefore give special attention to the realization of a robust and easy to operate experimental platform for the study of ultrafast nanoscale dynamics. The method we chose for looking at single nanoparticles is confocal microscopy, which is a widespread and powerful technique.^{45,46} All the experiments described in this review were based on the combination between confocal microscopes and ultrashort laser pulses. For all experiments, the appropriate laser beam was coupled into an inverted microscope and focused on the sample with an oil-immersion high numerical aperture (NA) microscope objective. The samples were raster-scanned using a piezoelectric scanner. The generated second harmonic, two-photon photoluminescence or fluorescence signals was collected in epi-confocal geometry and detected with a single-photon counting device for intensity imaging or electron multiplying charge coupled device (EMCCD) camera for spectral detection. For a detailed description of the experimental methods, we refer to original publications. The experiments involving phases and coherences were performed with the use of pulse shapers as active elements for controlling the phase of the laser fields.

1. INCOHERENT DYNAMICS IN A SINGLE MOLECULE

Our endeavors in studying ultrafast dynamics in individual particles began in 2005 when we adapted the ultrafast transient pump–probe technique, commonly used for ensemble measurements, to address for the first time the incoherent dynamics in single molecules at ambient conditions.^{12,26} To this end, we used laser pulses with a time duration of about 280 fs, derived from a Ti:sapphire oscillator and pulse-picked to a repetition rate of 1 MHz. The laser beam was propagated through a Michelson interferometer (see Figure 2a) and split into two parts. The outgoing laser pulse-pair with a mutual delay time Δt irradiated perylene- or cyanine-based molecule (PDI and DiD, respectively) in a confocal microscope. The first electronic transition in the molecule was excited to its saturation to gain access to the excited state energy redistribution dynamics. The experiment was only sensitive to incoherent dynamics for two reasons: the pulses were not short enough to access the coherent regime, and the lack of phase stabilization in the interferometer prevented the experiment from accessing phase information. The intensity of the laser was set such that a single excitation pulse drove the target transition to saturation, meaning that there was the same 50% probability for the molecule to be promoted to an excited state or to be stimulated back to the ground state by the same pulse. Assuming that the quantum efficiency for the studied molecules is close to unity this implies that the chance of emitting a fluorescent photon was also 50%. The presence of the second, probe pulse at the same time ($\Delta t = 0$) would not increase the probability for the molecule to end up in the excited state. Also, if the molecule would remain in the same excited vibronic state (or superposition of states) for a long time, $\tau \gg \Delta t$, the interaction with the second pulse would not affect the chance to excite the molecule to the excited state. In this case, the fluorescence signal would remain constant as a function of the interpulse delay time Δt . This balance, however, is broken when the lifetime of the excited state is short, that is, hundreds of femtoseconds to a few picoseconds. If the energy gets

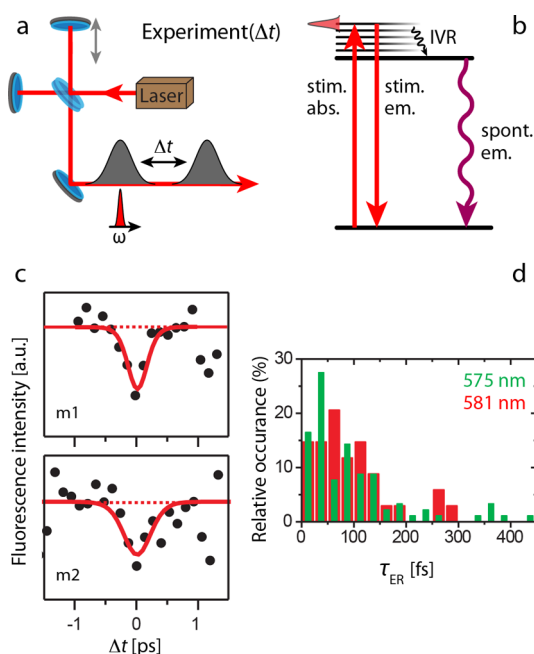


Figure 2. (a) Schematic of the *incoherent single-color* pump–probe experimental setup. (b) Concept of the experiment: interpulse delay time-dependent fluorescence signal provides information on the intramolecular vibrational relaxation (IVR) dynamics. (c) Typical fluorescence pump–probe traces for two individual molecules m1 and m2. (d) Histogram of the energy redistribution times τ_{ER} in PDI molecules for two different excitation wavelengths.^{12,26}

redistributed among other vibrational states in the excited electronic manifold, the excited state population becomes out of resonance with the second laser pulse. Therefore, the probability for the second pulse to stimulate the molecule to the ground state decreases with a time scale of the initially excited state lifetime τ_{ER} and the molecule has a bigger chance to remain in the excited state. Consequently, the probability of emitting spontaneous photons increases (up to a maximum of 75%). This is reflected in the modulation of the measured fluorescence signal as a function of Δt : at $\Delta t = 0$, the fluorescence signal is smaller than it is at longer delay times. The onset of the redistribution dynamics of the initially excited vibrational modes to other vibrational modes is directly encoded in the change (around $\Delta t = 0$) of the interpulse delay time-dependent fluorescence traces, examples of which are shown in Figure 2c. The red line is a result of the convolution of the energy redistribution time τ_{ER} with the autocorrelation of the excitation pulse.

The observed ultrafast energy redistribution may in principle involve two processes: the *intramolecular* vibrational relaxation (IVR)⁴³ and *intermolecular* energy transfer between the fluorophore and the encapsulating polymer molecules.⁴⁷ Structural and orientational diffusion are ignored here as they typically occur on a much longer time scale (few ps).⁴⁸ The extracted energy redistribution dynamics for the two investigated molecules (PDI and DiD) in the same polymer matrix were different. On the other hand, encapsulating DiD molecule in two different polymer matrices (PMMA or Zeonex) had only a minor effect on the extracted dynamics. Based on these findings we determined that it is the *intramolecular* interstate relaxation that determines the observed dynamics and that the intermolecular coupling is of less importance here. Interestingly we found that the extracted

relaxation times are independent of the exact excitation wavelength for both molecules (DiD and PDI) under study (see Figure 2d). This implies that on average the same relaxation pathways were probed for individual molecules. The large distribution of the extracted relaxation rates (~ 20 –400 fs) indicates a significant heterogeneity in conformation and local environment and consequently the intramolecular vibrational mode coupling among individual molecules.

The discussed pump–probe scheme can be extended from a single color scheme to a two-color scheme where the probe beam is Stokes-shifted and spectrally overlaps with the emission spectrum of the molecule.²⁷ In this case, the probe (or “dump”) beam can stimulate the excited molecules down to the ground state, leading to a decrease (depletion) in the spontaneous fluorescence signal, a process that underlies STED microscopy.^{49,50} Since the dump pulse can stimulate down only when the molecule is in the excited state, the fluorescence depletion dynamics reflects the excited state population lifetime. However, appreciable excited state depletion was only obtained for rather long, stretched dump pulses (~ 4 ps) and, thus, the experiment missed out on any dynamics occurring on a faster time-scale.

These experiments demonstrated that it is possible to probe the ultrafast dynamics in individual molecules at ambient conditions using an incoherent pump–probe technique in combination with confocal detection of the fluorescence signal. However, as already noticed, several limitations are inherent to this experimental approach. First, the finite spectral bandwidth of the laser field limits the temporal resolution of the experiment. It should be noted, however, that the use of much shorter pulses implies the loss of specificity as to which vibrational states are excited (because the broader spectral width of the pulse leads to excitation of multiple vibrational levels and averaging of state-specific dynamics). Second, the lack of a fixed phase relation between the pump and probe pulses prevents accessing coherent effects, which for large molecules at ambient conditions occur typically on a few 10s of fs.⁵¹ Finally, exciting the molecule in saturation leads to fast photobleaching, hence, limiting the measurement time.

The next logical step is to set up an experiment with the capability of addressing the coherent regime in the study of single molecules and nanoparticles. This essentially means that shorter pulses with active phase control need to be used.

2. PHASE CONTROL OF FEMTOSECOND PULSES

A necessary prerequisite for the study of coherent ultrafast dynamics at the nanoscale is to have precise control of ultrashort pulses with a spatial resolution comparable to the size of the nano-objects under study. Two fundamental problems arise when trying to achieve such a control in the focus of a confocal microscope.

- Ultrashort broadband pulses suffer from large phase/time distortions when propagating through optical components, in particular high numerical aperture (NA) microscope objectives.⁵² Quantifying and compensating for the phase distortions is paramount.
- The spatial resolution achievable with confocal microscopes is intrinsically limited by diffraction, whereas molecules and single nano-objects are subwavelength in size. Beating the diffraction limit is necessary to obtain nanoscale control of ultrashort pulses.

In a recent work we experimentally addressed these difficulties and demonstrated a general method that enables pulse control in a confocal microscope with high spatial and temporal resolution.³⁴ The main idea of our approach is to use small nanoprobe of deep subwavelength dimensions to extract information on the laser spectral phase and to overcome in a single experiment all the obstacles inherent to ultrafast (coherent) microscopy.

The nanoprobe of the spectral phase were second harmonic (SH) nanoparticles (NPs) of different materials: barium titanate (BaTiO_3), iron iodate ($\text{Fe(IO}_3)_3$), and gold (Au). When selecting the NPs for this particular application, it is strictly necessary that the particles do not present any resonance at the laser frequency, so they behave as probes, without further affecting the exciting spectral phase. Second harmonic generation (SHG) is widely used for femtosecond pulse characterization in particular with frequency-resolved optical grating (FROG);⁵³ spectral phase interferometry for direct electric-field reconstruction of ultrashort optical pulses (SPIDER);⁵⁴ and multiphoton intrapulse interference phase scan (MIIPS).⁵⁵ The main idea of these methods is that SHG, being a nonlinear coherent process, is sensitive to the spectral phase of the excitation electric field. This can be better understood with the aid of the sketch in Figure 3b. As a 15 fs laser pulse contains a broad range of frequencies, there are several different pathways that can lead to the same final field at the frequency 2ω . The different fields can interfere together to generate more or less amplitude at the frequency 2ω . Most efficient SHG occurs when all these fields are in phase.

In ref 56, a nanoversion of FROG was demonstrated, while in our work³⁴ we used MIIPS on SH NPs to demonstrate full pulse control on the nanoscale. In the experiment (see Figure 3a) we used a broadband Ti:sapphire oscillator providing pulses centered at 800 nm with a bandwidth of around 100 nm, at a repetition rate of 85 MHz. The laser beam was propagated through a pulse shaper based on a liquid-crystal spatial light modulator (SLM) and organized in a 4f configuration.^{57,58}

We first acquired SH confocal images of a sample containing BaTiO_3 NPs (Figure 3d) from which the position of the NPs could be inferred. Subsequently, we focused on a single SH NP of mean size of 180 nm and performed MIIPS on the nanoscale. In a MIIPS experiment⁵⁵ a series of known phase functions are imposed on the laser field using the SLM and the corresponding SH spectra are measured.

Because the phase is varied in a systematic way and the full SH spectral information is acquired, a single MIIPS iteration allows one to measure the spectral phase distortions at the sample position. The SLM is then used to compensate for these distortions and compress the pulse in time. In the case of our experiment,³⁴ this resulted in more than 1 order of magnitude increase in the SH intensity, which allowed imaging even smaller NPs (Figure 3e, red arrow), demonstrating great reproducibility and independence of the specific particle used.

We independently confirmed the accomplishment of pulse compression on the nanoscale by measuring interferometric autocorrelation traces (Figure 3h), from which we inferred pulse durations as short as 17 fs. Because in a typical pulse

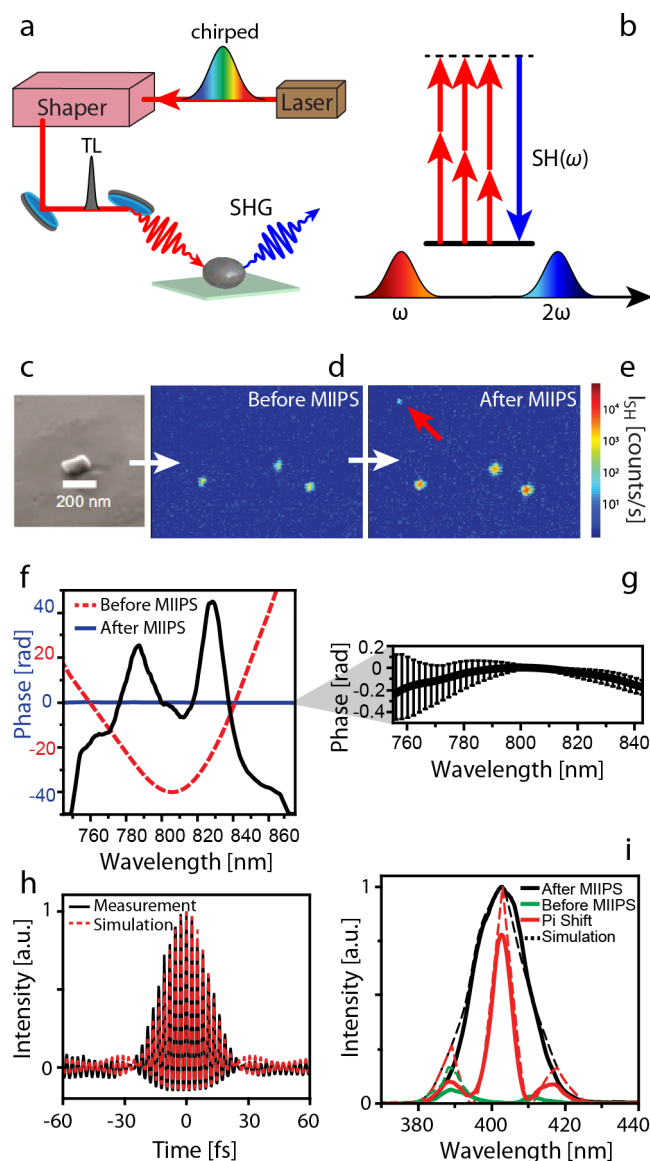


Figure 3. (a) Schematics of the pulse compression experiment on the nanoscale. (b) SHG from a broadband laser involving several different pathways. (c) Scanning electron microscope image of a BaTiO_3 NP. (d and e) SH images of the same sample area before and after pulse compression, respectively. (f) Laser spectrum (black) and the spectral phase before (red dashed) and after (blue) pulse compression. (g) Mean spectral phase resulting from 15 individual NP measurements. (h) Experimental interferometric autocorrelation of the laser pulses after compression (red dashed) and the simulated one (black). (i) SH spectrum before and after compression and with a π shift in the spectral phase applied at the center of the laser spectrum (solid lines) and the corresponding simulated spectra (dashed lines).³⁴

shaping experiment one also desires to control and manipulate at will the spectral phase of the pulse, in the same experimental work we applied a set of spectral phases on the SLM and compared the resulting SH spectra from individual NPs to the simulated ones. In all cases we found good agreement between theory and nanoscale experiment (Figure 3i).

These results demonstrate our ability to precisely control ultrashort laser pulses, in the phase and time domain, in the focus of a confocal microscope and with high spatial resolution. At the same time, they provide an easy to use method for

getting started with the investigation of ultrafast dynamics of single nano-objects in the coherent regime.

3. INTERMEZZO 1: PULSE CONTROL HURDLES

Let us take a short detour here. As already mentioned, the investigation of ultrafast (coherent) dynamics involves sending a tailored sequence of ultrashort pulses to a certain area of the sample and recording a certain observable while changing the experimental parameters, such as the interpulse delay or the laser spectral phase. In the cases described in the rest of this review, these experimental parameters are always controlled through the pulse shaper. It is then crucial that the pulse shaper itself does not introduce any additional experimental side effects.

In one of our previous works we showed that the degree of control of ultrashort pulses on the nanoscale can be hindered by the presence of spatiotemporal coupling.²⁸ Shaping of ultrashort laser pulses is typically done by dispersing the beam with a diffraction grating and then acting on each color separately using, for instance, the SLM. The inability to precisely recombine all the colors may lead to experimental artifact, which we generally refer to as spatiotemporal coupling.^{40,57,59,60} The problem with these kinds of artifacts is that they heavily depend on the specific configuration of the SLM. A reconfiguration of the SLM, as the one required to change the laser spectral phase or the interpulse delay, in general, produces different laser beams at the output of the pulse shaper. In order to keep spatiotemporal coupling under control, a method to check for these effects in the focus of a high NA objective is required.

In ref 28, we used gold nanoparticles with a diameter of ~ 100 nm to probe the spatiotemporal coupling in the focus of a confocal microscope. As the size of the NP was smaller than the diffraction limit, scanning the NP across the focal profile and performing simple single color pump–probe experiments allowed us to determine the extent of spatiotemporal coupling. Depending on the position of the NP within the focal spot, the photoluminescence signal from the gold NP showed strong variation as a function of the interpulse delay, for the shaper configuration used. This was a consequence of a variation of the focal width and changes in the symmetry of the focal beam profile with the interpulse delay Δt .

Fortunately, the observed spatiotemporal distortions could be largely eliminated by adapting the configuration of the shaper.⁶¹ Instead of out-coupling the beam after two passes through the active element, the beam is reflected back into the shaper so effectively, it passes through an active element four times. We reconfigured our shaper and tested this approach at even higher spatial resolution (few nanometers) by using individual DNQDI molecules. We scanned the molecule across the focal spot and recorded fluorescence intensity traces as a function of interpulse time delay. We found that placing the molecule at various positions within the focal spot yields exactly the same modulation in the fluorescence signal, indicative of fully homogeneous focal profile irrespective of the applied interpulse delay.

It is evident that, by using molecules, the pulse focus can be investigated with regard to spatiotemporal coupling. However, the use of small nonresonant SH NPs is more efficient as at the same time it enables one to compress femtosecond pulses to their Fourier-transform limit, control the spectral phase of laser pulses with high fidelity and characterize the focal spot of the

laser beam, all crucial requirements for the well-controlled ultrafast nanoscopy experiment.

It is worth mentioning here that the use of small NPs is also beneficial for characterization of the aberration of the optical elements, in particular the microscope objective. In ultrafast pump–probe experiments, either single or double color broadband pulses are used with spectral bandwidths of tens to hundreds of nm. It is important to ensure that all the wavelengths overlap in the same focal volume to ensure efficient interaction with the system under study. Microscope objectives are known to suffer from chromatic aberration, in particular, outside the most commonly used spectral range of approximately 500–650 nm. Detecting scattered laser light or SH from small NPs can be used to characterize the focal volume in terms of chromatic aberration. Filtering spectrally narrow bands out of the broadband pulse and recording the scattered intensity/SH in a confocal manner while scanning the NPs along the propagation axis provides a direct check on whether all the colors overlap in the same focal volume.

4. PLASMONIC ANTENNAS FOR ULTRAFAST NANOPHOTONICS

In the previous section we showed that the use of nonresonant NPs of dimensions smaller than the diffraction limit gives us full control over the spectral phase and, therefore, the time profile of a broadband laser pulse with an unprecedented spatial resolution. The logical successive step is then to apply our nanoscale pulse-shaping scheme to resonant NPs, which do have a coherent response.

Plasmonic nanoantennas are particularly promising in this sense as their localized surface plasmon resonances (LSPRs) can be engineered to overlap with laser fields in the visible and near-infrared region of the spectrum,^{62,63} as shown in the sketches in Figure 4b–d. An important characteristic of nanoantennas is their capability to confine a resonant propagating electromagnetic field in a small volume (hotspot) in the near field of the antenna. Combining this property of nanoantennas with ultrashort phase controlled laser pulses, the possibility of achieving coherent control of nanoscale plasmonic field was first theoretically predicted.⁶⁴ Experimental demonstrations of similar concepts, although neglecting the coherent response of the nanostructures, were also realized.^{13,14} Many studies determined the coherence time of surface plasmons in resonant nanoantennas to be in the 15 fs range and therefore at the limit of the time resolution achievable by broadband laser systems.^{20,65–68}

In our group we recently carried out a series of experiments aimed at detecting, understanding and making use of the ultrafast resonant response of plasmonic nanoantennas.^{33,35,36} These experiments rely on the detection of nonlinear optical signals from the nanoantennas, such as SHG and two-photon induced photoluminescence (TPPL). When a broadband laser field impinges on a resonant nanoantenna, the interaction is most efficient for the laser frequencies that overlap with the LSPR and which are efficiently localized in the hotspot of the nanoantenna. The localized field is therefore a function of both the incoming field and the LSPR. As a consequence, SHG and TPPL, generated in the proximity of the nanoantenna, are both affected by the resonance response (Figure 4c,d).

By observing such nonlinear signals, we first demonstrated that the resonant response of nanoantennas could be measured in its phase and amplitude components.^{33,35,36} This is important, as the possibility of correctly exploiting LSPRs is

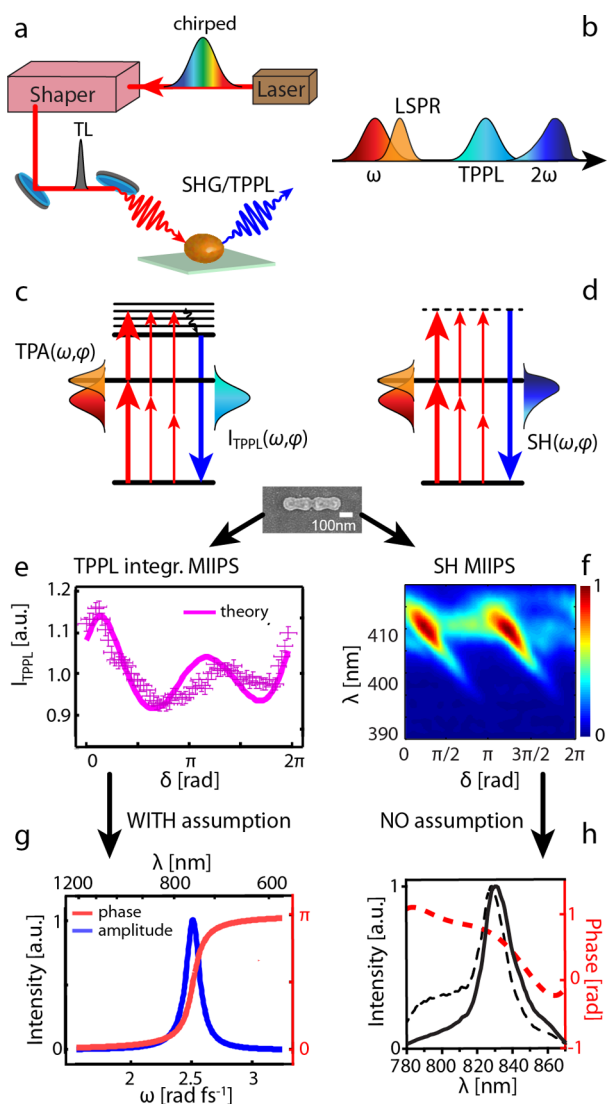


Figure 4. (a) Schematics of the ultrafast nonlinear experiments on plasmonic nanoantennas. (b) Frequency domain representation of a LSPR at the fundamental laser frequency and the TPPL and SHG from a nanoantenna. (c) TPA and TPPL processes in a resonant nanoantenna involving several pathways, with relative importance weighted by the LSPR (thick red arrows). (d) SH processes in a resonant nanoantenna. (e) TPPL intensity MIIPS trace measured on a single nanoantenna and fit to the data based on a Lorentzian response. (f) Spectrally resolved SH MIIPS trace measured on a single nanoantenna (g) Lorentzian phase and amplitude response retrieved from the fit of the TPPL MIIPS trace. (h) Phase and amplitude response and comparison to the laser spectrum, as obtained from the SH MIIPS trace, free of additional assumptions.^{33,36}

subject to the precise understanding of the effect of the resonances on the incoming laser field. In the experiments we used the nanoscale MIIPS method described above and applied it to individual nanoantennas, detecting either the TPPL intensity³³ or the SH spectrum^{35,36} (Figure 4e,f). A fit of the MIIPS traces allows the complete (amplitude and phase) resonant response to be retrieved (Figure 4g,h). Both experimental approaches present advantages and limitations, and the choice of using one or the other should depend on the specific experimental requirement. The main advantage of using the TPPL signal is that it is generally larger and, thus, more easily detectable than the SH from individual nanoantennas,

especially considering that for spectral measurements the SH is divided between several pixels of the EM-CCD camera. However, the TPPL spectrum, unlike the SH, is independent of the laser spectral phase and only its intensity varies as a function of the applied phase. A considerable amount of information contained in a spectrally resolved SH MIIPS trace is therefore lost when detecting the TPPL. As a consequence, an assumption about the shape of the LSPR is necessary to extract information from a TPPL MIIPS trace. In the case of ref 33 we made the assumption that the nanoantenna response could be well approximated by a Lorentzian function (Figure 4g), for which both the phase and the amplitude components are different functions of the same two parameters: the central frequency and the width of the Lorentzian. In contrast, in the case of SH MIIPS, the full information is contained in the spectrally resolved traces, and no additional assumption is needed. In ref 36 we showed that, in the case of simple nanorod antennas, the results obtained were compatible with a Lorentzian model, confirming the agreement between the two methods for this simple case. Nevertheless, for an arbitrary and more complex plasmonic structure, for which an accurate prediction on the shape of the resonance is not straightforward, the SH MIIPS approach is a more powerful method to extract information about the LSPR.

These results demonstrate that, by using ultrashort phase-controlled pulses, together with a reliable characterization method, the effect of LSPRs in nanoantennas can be measured. In order to actively exploit these LSPRs, in refs 33, 35, and 36 we compared the responses of different nanoantennas resonant with opposite sides of the laser spectrum, subject to precise phase shaping of the laser pulse. We demonstrated two different applications: an ultrafast plasmonic switch³³ (Figure 5a,b) and a multicolor SH imaging technique³⁵ (Figure 5c,d).

In the first experiment, based on the theoretical proposal by Stockman and collaborators,⁶⁴ we used a pair of coupled nanorods of different lengths (respectively resonant on the blue and red side of the laser spectrum) to move the localized energy from one side to the other of the plasmonic nanostructure on an ultrafast time scale. By exciting the system with a chirped pulse, in which the blue colors arrive at the sample before the red ones, the two different nanorods were separately addressed, with a switching time of ~ 100 fs. As shown in Figure 5a,b, the light localization was probed with a second Fourier limited pulse and the TPPL was used as the experimental observable. Correctly designed plasmonic structures combined with the ultrashort laser pulses used to excite them have the potential to generate switchable hotspots with high temporal resolution and highly confined energy. This idea can be exploited to selectively excite single molecules at fs time scale within a close packed ensemble using the nanoantennas.

The second application, illustrated in Figure 5c,d is a multicolor SH imaging technique.³⁵ Because the SH spectra from nanoantennas depend on the LSPRs, different nanoantennas in general emit different SH spectra. In a sample that contains two different species of nanoantennas this gives the opportunity to distinguish them based on their SH spectra. This has important implications in biological imaging as it allows one to label different components of a tissue using different nanoantennas; excite them all at the same time with the same ultrafast laser and then use spectrally selective detection at the SH wavelength to discriminate between the different nanoantennas. A similar experimental technique, often named multicolor imaging, is used regularly to image different

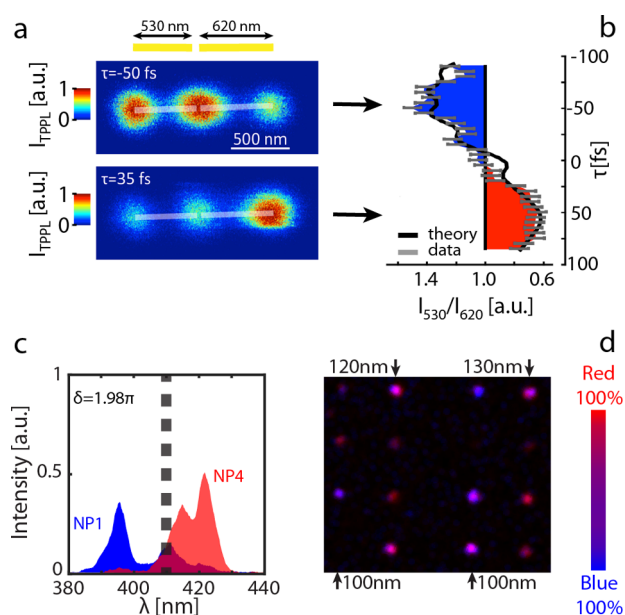


Figure 5. Hotspot switching and multicolor imaging. (a) TPPL image of two coupled nanoantennas excited by a chirped pulse and probed by a transform-limited pulse at two interpulse time delays. (b) The luminescent hotspot switches on fs time scale as the chirp runs over the coupled antenna. (c) SH spectral response of two distinct nanoantennas excited by a broadband laser pulse with the spectral phase that maximizes the spectral contrast. (d) Two-color SH image of an array of nanorods: change in the coloring of different nanorods allows us to readily distinguish the rods of different lengths.^{33,35}

fluorescent markers, such as semiconductor QDs, but prior to our work³⁵ had never been extended to the SH range. In recent works, the possibility of using nonresonant SH NPs, such as BaTiO₃ NPs, as markers for SH imaging of tissues was considered.^{69,70} Benefits of SH imaging include high stability and absence of photo blinking and bleaching; relatively narrowband emission, allowing for easy spectral separation from the sample autofluorescence; use of near-infrared excitation, which limits photodamage of the sample and increases the penetration depth for deep tissue imaging. In our work³⁵ we showed that, by using resonant nanoantennas together with broadband phase controlled laser pulses, multicolor imaging in the SH range can be achieved. Figure 5c shows SH spectra from two different nanoantennas, acquired with a specifically tailored spectral phase that maximizes their relative spectral difference. Sending the blue and the red part of the SH (splitting at 415 nm) to different APDs allowed us to discriminate between the two nanoantenna species, as shown in Figure 5d, which demonstrates the achievement of multicolor SH imaging.

So far we have presented and discussed a toolbox for performing phase-controlled ultrafast experiments on the nanoscale. Plasmonic nanoantennas have provided a good platform to test not only our highly sensitive phase measurement scheme, but also our ability to actively control nanoscale responses. Clearly, these concepts are equally suitable to address single molecule dynamics in the coherent regime. Single molecules under ultrafast phase-controlled excitation are the focus of the second part of this review. The final objective would be to combine molecular and plasmonic systems together and still be able to perform ultrafast coherent control.

We will briefly touch on one experiment on single molecules and nanoantennas in the last section of this review.

In the following we demonstrate that having a precise control over the time delay Δt and relative phase $\Delta\phi$ between the pulses allows the study of coherent properties of individual molecules and molecular complexes.

In the incoherent pump–probe experiments discussed in the first section, we operated in the excitation saturation regime. Hence, in these experiments, the interaction of a molecule with the laser light produced an excited state population, and excited state population dynamics was reported. Now, in all of the following, rather than limit ourselves to population, we switch to quantum mechanical syntax and address transition probabilities. In these experiments we operate in a weak excitation regime and we exploit quantum coherence, that is, a state where a single pulse creates a coherent superposition of two or more states in a molecule. Interaction with the second, delayed pulse probes the phase memory of that coherent superposition in time. After the interaction of the molecule with the pulse-pair, the probability for the molecule to be in the excited state becomes a function of both the interpulse delay time and the phase difference. This leads to a modulation of the excitation probability, which is reflected in a modulation of the fluorescence signal.

One of the holy grails in physical chemistry is the ability to control the behavior of a molecular system. In spectroscopy this implies that the interaction of a molecule with light leads to a localization of the energy in a specific predetermined mode, specific energy redistribution pathway, or/and specific reactivity pathway. In the experiments presented below, the detection and control aspects of the coherence are interrelated and indistinguishable. In fact, the detection of the coherence is realized based on the fact that the population probability depends on the interpulse phase relation.

5. DETECTION AND CONTROL OF FEMTOSECOND COHERENCE IN SINGLE MOLECULE

In a simple two-level system, the interaction with a coherent laser field creates quantum coherence between the electronic ground and excited states. This coherence leads to Rabi oscillations, which in simple terms is a periodic change (with the so-called Rabi frequency) of the probability for the molecule to be in the ground or excited state. Rabi oscillations in individual molecules and quantum dots have been detected recently at cryogenic conditions.^{71,72}

In our experiment we explored and controlled the coherence in individual terrylenediimide (TDI) molecules at ambient conditions.³² TDI molecules were resonantly excited at 630 nm with a phase-locked pulse-pair derived from an optical parametric oscillator and the resulting fluorescence was detected in a confocal manner. A pulse picker was used to reduce the repetition rate from 76 MHz to 500 kHz, allowing higher peak power while still providing long observation time of a single molecule before bleaching. The pulses were compressed to their Fourier limit (pulse width of ~ 75 fs) and, which is crucial for the success of the experiment, the interpulse phase relation was precisely controlled using an acousto-optic programmable dispersive filter (AOPDF)-based pulse shaper.

Typical interpulse delay dependent fluorescence traces from three individual molecules with $\Delta\phi$ fixed at 0 rad are shown in Figure 6c. The three traces exhibit variation in the fluorescence. The fluorescence counts decrease or increase as a function of the interpulse delay time Δt within several tens of femto-

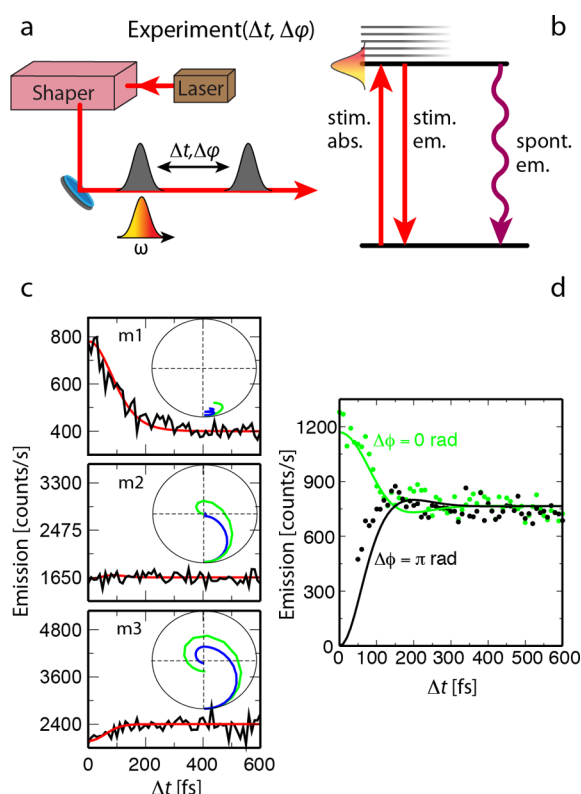


Figure 6. (a) Schematic of the *coherent quasi-broadband pump–probe* experimental setup. (b) Concept of the experiment: an excitation pulse creates a coherent superposition of the electronic ground and excited states, which is subsequently probed with a second pulse. (c) Coherent properties of the molecule are reflected in the interpulse delay time-dependent fluorescence traces, providing information on the trajectory of the Bloch vector: green, 0 fs time delay; blue, 600 fs delay. (d) π -Flip between the two pulses does invert the interpulse delay time-dependent fluorescence traces.³²

seconds, after which they remain constant. The observed dynamics reflects the decay of the coherence between the ground and the excited electronic states, with a time constant of about 50 fs. The effects of the incoherent excited state relaxation processes are excluded here as they typically occur on much longer time scales (hundreds of fs to ps).⁷³ The observed turnover from decaying to flat and growing character of the fluorescence trace (top to bottom in Figure 6c) is attributed to an increased strength of interaction between the transition dipole moment of the molecule and the laser pulse. In other words the laser field drives the transition more effectively and the Rabi frequency increases. The red lines in Figure 6c represent numerical simulations based on the optical Bloch equations.

The state of a two level system is often represented in the form of a Bloch vector and its evolution is given by the time-dependent trajectories of the vector tip on the Bloch sphere, as shown in the insets in Figure 6c. In the case of molecule m1 (Figure 6c, top), the Rabi frequency is small and mainly absorption takes place, while very little stimulated emission. Due to the rapid dephasing, the Bloch vector becomes shorter and the probability for excited state population as a function of Δt decreases. As the interaction between the molecule and the laser field increases (higher Rabi frequency), the trajectories become longer and flip around the Bloch sphere. As a result, the dephasing at larger Δt can leave a higher excited state

probability, as in the case of molecules m2 and m3 in Figure 6c. Due to the finite pulse duration (approximately 75 fs) and short dephasing time (~ 50 fs), complete Rabi oscillation could not be resolved in our data. However, from the calculations (red lines in Figure 6c), we determined the periods of Rabi oscillations ranging from 20 to 100 fs.

Precise control over interpulse phase relation allowed us to manipulate the coherent state of the molecule. The largest contrast was obtained for $\Delta\phi = \pi$, as π phase difference reverses the rotation direction of the Bloch vector, hence, increasing the probability for the molecule to end up in the opposite state, that is, the ground state. In Figure 6d we show two fluorescence traces as a function of the interpulse delay time Δt recorded on the same molecule for the interpulse phase $\Delta\phi = 0$ and $\Delta\phi = \pi$ rad. Within the coherence time (<100 fs) the in-phase pulse-pair interferes constructively and enhances the excited state population probability (higher fluorescence counts; black data points), whereas the out-of-phase pulse-pair interferes destructively leading to a decrease of the excited state population probability (lower fluorescence counts; green data points).

These experiments clearly demonstrate the ability to prepare a molecule in a specific state. The dephasing time at ambient conditions is very short (<100 fs) and imposes the necessity to use ultrashort pulses. The precision with which we can control the state of a molecule or, in other words, the achievable contrast (the molecule resides in the excited or ground state) depends mainly on temporal fluctuations of the interaction strength between the molecule and the laser field and the precision with which one controls the interpulse phase. The success of these experiments evidently lies in our ability to control intra- and interpulse phase with high finesse.

6. VIBRATIONAL WAVE PACKET DYNAMICS IN SINGLE MOLECULES

In the previous section we discussed the simplest manifestation of quantum coherence between the two electronic levels of a molecule. Other than preparing the molecule in an arbitrary superposition of the two states, we demonstrated the ability to enhance or decrease the probability for the molecule to be promoted to the excited state. Exploiting this coherence, however, does not provide any handles to deterministically steer a molecule to a specific final state out of a multitude of accessible states. It only allows tipping the scale of probability for the molecule to end up in one of the two states (ground or excited).

The next experiment addressed in this review focuses on visualizing and controlling coherence within a single electronic excited state but involving multiple vibrational states.³⁰ Analogous to previous experiments, also in this work we used a pair of femtosecond pulses with well-controlled interpulse delay time Δt and phase shift $\Delta\phi$. The laser pulses were derived from an ultrabroadband Ti:sapphire oscillator, operating at 85 MHz and delivering pulses with a bandwidth of 120 nm, centered at 680 nm. Pulse compression, generation of pulse-pair, and control of the interpulse phase were performed with a 4f-shaper based on a SLM.

As a molecule of study we used a rylene homologue called DNQDI.⁷⁴ Single molecules were illuminated with a pulse of sufficient bandwidth to excite several vibrational levels in the electronic excited state and thus generate a quantum wave packet.⁷⁵ The excited wave packet travels in time within the excited state potential back and forth until the phase relation

between individual vibrational states (coherence) is lost. The second, delayed, pulse leads to generation of a second wave packet. If the two wave packets are generated within the coherence time, quantum interference between them leads to an enhancement or suppression of the excited state population probability and, therefore, to modulation of the fluorescence intensity. The fluorescence traces as a function of the interpulse delay time Δt and fixed interpulse phase difference, for three exemplary molecules, are shown in Figure 7c. The traces are

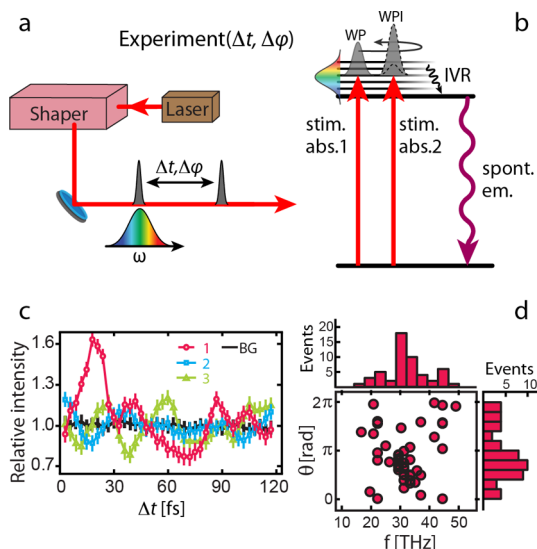


Figure 7. (a) Schematic of the *coherent broadband pump–probe* experimental setup. (b) Concept of the experiment: an excitation pulse creates a coherent superposition of a set of the vibrational states (vibrational wave packet, WP), which is subsequently probed through an interference with a wave packet created by the second pulse. The coherent properties of the vibrational wave packet interference are reflected in the interpulse delay time-dependent fluorescence traces.³⁰

normalized to the fluorescence intensity at long delay times. All traces exhibit oscillatory character with a typical modulation in the order of $\pm 10\%$ of the average fluorescence level. As explained earlier, these oscillations reflect the quantum wave packet interference. The oscillations with a period of 30–40 fs typically vanish on a time scale of 100 fs, indicative of the wave packet phase memory loss, that is, dephasing. Fourier analysis of the fluorescence traces revealed a single, dominant component with a frequency of around 33 THz (see Figure 7d). The extracted frequency of 33 THz (or 1000 cm^{-1}) reflects the travel time of the wave packet within the excited state potential and is therefore directly linked to the energy separation of the vibrational states comprising that wave packet. The variation in the starting phase of the oscillations among individual molecules is attributed to the large heterogeneity of their characteristic vibrational progression, that is, spectral positions, widths, and strengths.

The generation and, in particular, control of a vibrational wave packet gives much more control over the molecule than the interelectronic state coherence discussed in the previous section. We can shape the pulse in frequency domain to form a wave packet of arbitrary states. Coupling of these states within the vibrational manifold and with extramolecular modes gives plenty of possible energy relaxation and energy transfer pathways. Hence, optimization of the time-phase structure of the exciting pulse-pair can be used to excite individual

molecules at ambient conditions to a desired, specific target state.

7. INTERMEZZO 2 – INTERPULSE PHASE RELATION AND SINGLE MOLECULE EXCITATION SPECTROSCOPY

We have just learned that it is possible to launch and probe the evolution of a vibrational wave packet in a single molecule at ambient conditions provided that the laser pulses have sufficient bandwidth and that we can precisely control the interpulse phase relation. The use of the shaper allowed us to keep the relative phase between the pulses fixed at $\Delta\phi$ and scan only the interpulse time delay. By changing the phase relation from in-phase ($\Delta\phi = 0$) to out-of-phase ($\Delta\phi = \pi$) the phase of the fluorescence trace recorded from the same molecule changes by π as well, as shown in Figure 8a. This is a clear demonstration that by tuning the interpulse phase we can arbitrarily control the excited state population probability.

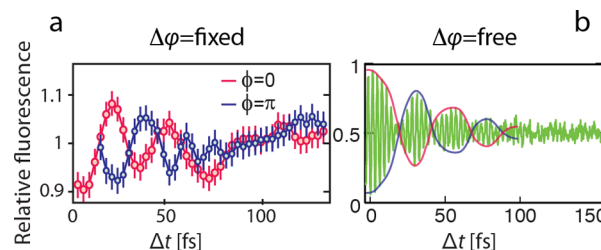


Figure 8. (a) Delay time-dependent fluorescence traces recorded from a single DNQDI molecule for a set interpulse phase difference of 0 (red) and π (blue). The observed oscillatory traces reflect the beating due to the vibrational sideband.³⁰ (b) Interferometric fluorescence trace as a function of the interpulse delay recorded from an individual QDI molecule.⁷⁶

An alternative approach is to let the interpulse phase $\Delta\phi$ change together with the interpulse delay time Δt . Recently we performed such an experiment on individual QDI molecules, where instead of phase-locked pulses, the pulse-pair was generated in a Michelson interferometer using a motorized delay line.⁷⁶ The outgoing pulses continuously interfere with each other as a function of the interpulse delay time Δt . The interfering pulse-pair excites the molecules, and the resulting fluorescence is detected as a function of the interpulse delay time Δt . Effectively, different excitation wavelengths are sampled when scanning a time delay between two interfering broadband (over 100 nm) laser pulses. The fluorescence traces exhibit clear modulation on at least two time scales (see Figure 8b). The fast oscillations with a period of ~ 2.3 fs (corresponding to the central wavelength of the broadband pulse of 700 nm) result from the interference of the carrier frequency of the two pulses and, superimposed, slower beating signal with a period of about 50 fs. Analogous to Fourier transform spectroscopy, in order to extract the effect of the molecule on the measured fluorescence traces (interferograms), we first recorded an interferogram of the laser itself, Fourier transformed both signals, and divided them in the frequency domain. This way we obtained the excitation spectrum of a single QDI molecule. The observed beating frequency in the time-dependent fluorescence traces comes from the beating of the two vibronic bands in the excitation spectrum of the molecule.

It should be realized here that, even though the two experiments are performed in a different manner, the phase-locked experiment requires a spectral shaper in order to lock the pulses at the central laser frequency, while the interferometric experiment is performed with arbitrary pulses (or CW excitation) and the interpulse phase is scanned; the obtained information on the vibrational states is equivalent.⁷⁷ Oscillations in the fluorescence signal in the phase-locked experiment reflect the frequency difference of the two beating vibrational bands. In the interferometric experiment we get the full excitation spectrum of the molecule, hence, implicitly the frequency difference between the vibrational states that were excited.

8. TWO-COLOR PHASE-CONTROLLED DETECTION OF ENERGY TRANSFER IN LIGHT HARVESTING COMPLEXES

In the previous sections we discussed experiments that uncover coherent properties in individual molecules: Rabi oscillations and vibrational wave packet interference. In nature, however, complex (molecular) architectures most of the time not only involve modes that are intricately coupled to each other but also involve energy transfer between molecular units. An example of such system is the light harvesting complex 2 (LH2), a biologically relevant macromolecule that is found in purple bacteria. LH2s are the main antenna complexes in the light harvesting process and as such they play a crucial role in photosynthesis. Coherent effects have been observed in LH2 complexes in bulk experiments.^{78–80} Consequently, it has been speculated that coherent delocalization of the excitation energy may be responsible for the observed high efficiency of the energy conversion in such photosynthetic organisms.

LH2 are complex architectures that, in simple terms, are built of two pigment–protein rings, which give rise to distinct electronic transitions called B800 and B850, according to the wavelengths at which they absorb light (see Figure 9).^{81,82} The energy absorbed by the B800 band is transferred to the B850 band and funneled toward the reaction center. In our experiment we addressed the coherent character of the B800–B850 energy transfer step in individual LH2 complexes.³¹ To this end we designed a two-color experiment in which the two transform-limited pulses are resonant with the B800 and B850 band, respectively. We detected the fluorescence signal as a function of the time delay Δt between the two pulses, keeping the relative phase between the pulses constant. The pulse-pair used was generated through phase-shaping in the pulse shaper. Evidently, the experiments discussed in earlier sections are of extreme importance here, as the success of this experiment is highly dependent on the control over spectral phase of the ultrashort pulses. Moreover the experiment has to be free of spatiotemporal coupling in order to probe solely excitation-pulse induced effects.

The measured time-dependent fluorescence traces recorded from individual molecules exhibit oscillations with a period of about 200 fs and lasting over 400 fs (exemplary trace is shown in Figure 9e). The observed oscillations are ascribed to quantum interference between two excitation paths that populate the same target state, that is the emitting lowest-energy B850 state. The blue-shifted pulse induces excitation in the B800 band, which in time repeatedly acquires character of the B850 band. In other words, the interband B800–B850 electronic coupling, possibly assisted by resonant low-energy vibrational modes, make the excited electrons travel back and

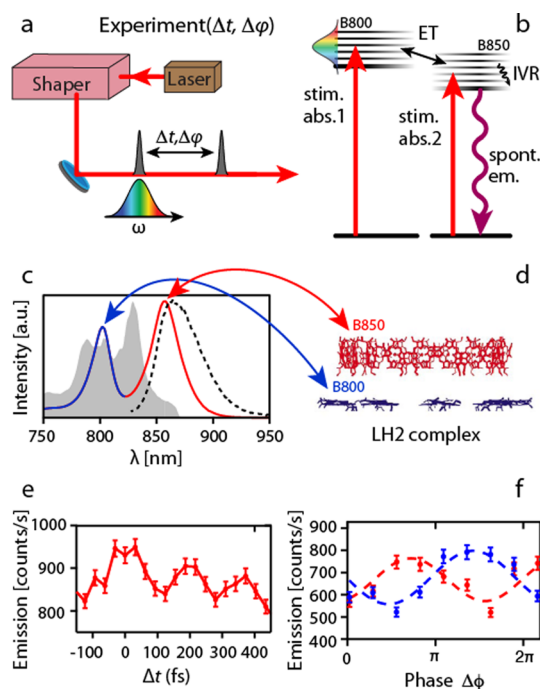


Figure 9. (a) Schematic of the *coherent broadband pump–probe* experimental setup. (b) Concept of the experiment: an excitation pulse creates an excited state population in the B800 band of the LH2 complex. The energy transfer to the B850 band is subsequently probed with a second pulse, resonant with the B850 band. The coherent properties of the intermolecular energy transfer are reflected in the interpulse delay time-dependent fluorescence traces. (c) Blue/red line represents the absorption spectrum of the LH2 ensemble solution. Gray, shaded area reflects the laser spectrum. Dashed, black line indicates emission spectrum of the LH2 ensemble solution. Green, solid line is the phase profile applied to the laser spectrum in order to generate a pulse-pair. (d) Simplified representation of the LH2 complex structure. Chromophores are organized in two rings giving rise to the distinct absorption bands, called B800 (blue) and B850 (red). (e) Exemplary interpulse delay time-dependent fluorescence trace recorded with a fixed phase difference between the pulses from a single LH2 complex. (f) Phase-sweep traces recorded on the same molecule for a fixed interpulse time delay. The red and blue trace result from averaging the first three and subsequent seven consecutive phase-sweep traces.³¹

forth within the coupled B800–B850 potential energy surface. The second, red-shifted pulse can then either enhance or reduce the population of the emitting B850 state through quantum interference, thus, giving the fluorescence trace its oscillatory character.

Probing the coherence in many individual LH2s we found a rather large spread of the oscillation periods, ranging from 100 to 400 fs. Given the complexity of LH2, the involvement of many modes and contribution from thermal fluctuations, it is understandable that slightly different energy transfer pathways will be optimal for individual complexes.

It is important to note here that the coherence in this particular energy transfer process on average persists much longer (>200 fs) than a typical coherence in isolated fluorophore in polymer matrix (~50 fs).^{32,44} Even though the intrinsic strength and typical time scale of the coherence differs among complexes, the robustness of the intermode coupling in combination with the extensive protective protein scaffold appears to effectively screen the two modes from coupling to other energy dissipation channels.

Finally, we attempted to influence the final fluorescence probability of a single LH2. Sweeping the interpulse phase relation between 0 and 2π at constant time delay between the two excitation pulses $\Delta t = 100$ fs lead to oscillatory traces indicating that we can externally influence the efficiency of populating the emitting B850 state (see Figure 9f). So here once again it is important to have full time and phase control over the laser pulses; otherwise, it is impossible to carry out such an experiment.

By repeating the phase sweep ($0-2\pi$) scan multiple times on the same complex we found that the phase of the measured oscillatory fluorescence signal changes occasionally on a time scale of some 10s of seconds (red and blue trace in Figure 9f). Apparently, even within the same complex different energy transfer pathways can dominate at different times.

In the experiments highlighted in the two previous sections, we probed and controlled the final excited state population in individual molecules. The experiment described here forms yet another step toward the ability of steering the chemical reactions at a single molecule level, as we are effectively able to control the energy transfer efficiency between spatially distinct multimolecular rings within one LH2 complex. These pioneering experiments pave the road for investigation of other ultrafast energy transfer and delocalization processes on the nanoscale.

9. INTERMEZZO 3: MOLECULES MEET PLASMONICS

The detection of single molecules and the investigation of molecular dynamics through fluorescence is still the most common choice in laboratories around the world as it typically delivers a superb signal-to-background ratio. However, in many cases, such as for the LH2 complexes, fast photobleaching, low fluorescence quantum efficiency and ultimately the limited fluorescence photon budget hinder progress in the investigation of ultrafast dynamics at a single molecule level at ambient conditions.

It has been demonstrated that plasmonic nanoantennas can boost the brightness of fluorophores by concentrating the electromagnetic fields into subdiffraction limited volumes.^{83,84} Coupling of the emitter (a molecule or a quantum dot) to a plasmonic nanoantenna leads to an increase of the excitation efficiency of the emitter and the modification of its radiative and nonradiative rates, which can increase the overall quantum yield of the emitters.⁸⁵

Recently we investigated the properties of LH2 complexes coupled to plasmonic antennas.⁸⁶ Even though the experiments are not time-resolved, in the light of this review it seems appropriate to discuss them since with these experiments we took the first steps toward studying ultrafast processes in coupled plasmonic-molecular systems.⁸⁶ Briefly, LH2 complexes were spin-coated on a microscope slide with gold nanorod antenna arrays, with a concentration that yielded only a few LH2 complexes per antenna. For optimal fluorescence enhancement both excitation enhancement and quantum efficiency (QE) enhancement are important. Therefore, the antenna length of 160 nm was chosen because its gold plasmon resonance overlaps well with both the absorption (800 nm) and emission band (900 nm) of the LH2. The molecules were excited at 800 nm with ~ 100 fs pulses in a confocal microscope and the resulting fluorescence detected with a single-photon counting device.

Confocal images of the LH2 complexes were recorded with and without the presence of the antenna array. Images were

then normalized to the excitation power and the fluorescence counts were directly compared. We found an over 500-fold fluorescence enhancement for LH2 complexes coupled to a gold nanoantennas. Further analysis revealed that the resonant antenna produces an excitation enhancement of roughly 90 times and a fluorescence lifetime shortening to about 20 ps (from the initial ~ 1 ns). At the same time, the radiative rate enhancement results in a 5.5-fold-improved fluorescence QE. The correlation between the fluorescence enhancement factor and the fluorescence lifetime for a number of investigated molecules is shown in Figure 10b. For the $>500\times$ enhanced

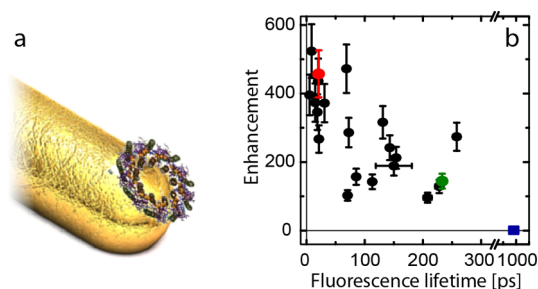


Figure 10. (a) Artist impression of a single LH2 complex coupled to a gold plasmonic antenna at the location of the plasmonic hotspot. (b) Enhancement factor as a function of fluorescence lifetime for a number of investigated individual LH2 complexes; blue reference point for free LH2 complex without plasmonic antenna.

LH2 complex, the total count rate per molecule was $15\times$ increased compared to the unenhanced case. This significant increase in the total fluorescence photon budget allowed us to record photon antibunching of a single light-harvesting complex under ambient conditions, revealing that this multi-chromophoric complex behaves as a nonclassical single-photon emitter.

Evidently the presented approach of coupling molecules to plasmonic structures holds great promise toward studying weakly fluorescent, natural molecular systems. In particular, it enables further, detailed study of the energy transfer processes and the role of quantum coherence at the level of single complexes.

CONCLUSIONS AND OUTLOOK

We have shown that a broadband Fourier-limited pulse focused in a diffraction-limited spot is a perfect basis to probe and control individual nano-objects such as plasmonic nanoantennas and individual molecular systems. Addressing plasmonic resonances, their spectral phase response is retrieved and exploited to control the spectral response or produce dynamical nanoscale hotspots. Turning to molecular systems, the whole toolbox of coherent control is brought to single units, allowing us to prepare coherent states and trace coherent energy transfer even of complex molecular systems in a highly inhomogeneous environment.

In all these experiments, control and stability of the spectral phase of the pulse has turned out to be crucially important. Measuring coherent effects in energy transfer processes or intrinsic coherences within vibrational wave-packets, the complexity of the system and its environment produce a large heterogeneity in the extracted time scales and magnitudes. Similarly for plasmonic structures, despite great advances in nanofabrication, no two plasmonic structures are identical, leading to distinct heterogeneity in coherent response between

these structures. Therefore, pulses have to be controlled with high precision in order to probe the inhomogeneities between individual nanoparticles and not the uncertainty in spectral phase of the pulse.

In some of the presented experiments, to keep control and avoid artifacts, we probed the coherences using pulses with flat spectral phase and intensity given by the laser spectrum. Alternatively, one could shape the laser pulse in terms of spectral intensity and phase relation to address optimally specific levels and influence chosen processes. Ultimately, one could aim for closed loop coherent control on a single unit, which is quite challenging in the case of molecules emitting only a limited number of photons before photobleaching.⁷⁴

We have shown that one can prepare the molecule in specific coherent state and probe its coherent energy transfer and relaxation pathways. It would be truly groundbreaking if the two experiments could be combined together in a three-pulse experiment: preparing a certain designed superposition of states, and next, manipulate its temporal evolution toward the anticipated final state.

Detailed knowledge of the spatiotemporal coherent response of plasmonic structures is essential to design more complex plasmonic architectures capable of localizing electric fields in a nanovolume with tailored temporal and phase character. Such design brings us a step closer toward the realization of pump–probe schemes on the nanoscale. For ultrafast nanoscopy experiments on individual molecules the sample concentration is not so critical, as the local enhancement in the engineered nanovolume will dominate the detected signal. For more complex and dense molecular samples, such as bacterial membranes in which spatial coherent energy transfer among various complexes is expected, the perspective to exert a coherent pump–probe experiment on a chosen complex, or between two complexes, is truly fascinating.

Unfortunately, all the presented experiments suffer from the fundamentally limited photostability of individual fluorescent molecules. Moreover some experimental concepts require saturating excitation conditions, while some molecules are intrinsically designed for energy transfer and avoid spontaneous emission (LH2). In spite of these limits, the detection of fluorescence is still winning over alternative detection schemes in terms of simplicity of the experiment and the achievable signal-to-background ratio. Also, while relying on an incoherent detection scheme, we have demonstrated that the coherent properties of individual nanoparticles can be successfully accessed by dedicated control of the excitation path. The limitations of the presented experiments can be partially lifted by coupling molecular systems to plasmonic antennas, which speed up the radiative decay rate, resulting in a higher quantum efficiency and photon stream, combined with reduced bleaching and higher total number of photons emitted. Still, detection of a coherent response would be preferred, such as absorption, stimulated emission, or four-wave mixing, yet detection of single units along these routes is equally challenging.

Clearly ultrafast nanophotonics is a field in action. This review provides a timely snapshot, and surely, significant advances can be expected in the near future.

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Notes

The authors declare no competing financial interest.

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